

THE SYNTHESIS AND ANTI-MRSA ACTIVITY OF AMIDINIUM-SUBSTITUTED 2-DIBENZOFURANYLCARBAPENEMS

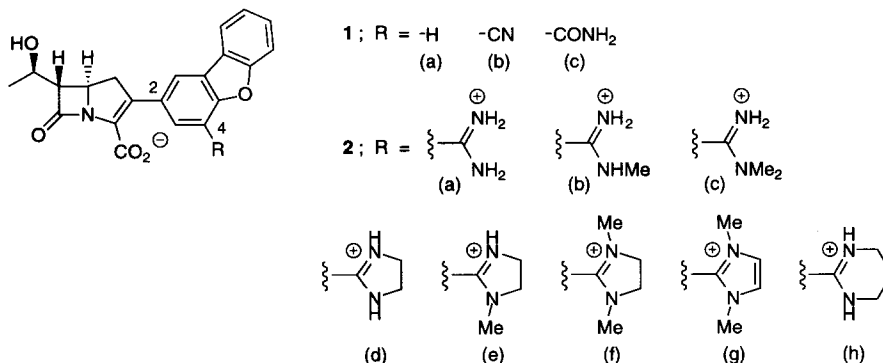
Joanne B. Laub,^a Mark L. Greenlee,^{a*} Frank DiNinno,^a
Joann L. Huber,^b and Jon G. Sundelof^b

Departments of ^aMedicinal Chemistry and ^bInfectious Disease
Merck Research Laboratories, P.O. Box 2000, Rahway, NJ 07065, U.S.A.

Received 20 August 1999; accepted 8 September 1999

Abstract: A series of amidinium-substituted 2-dibenzofuranylcabapenems with potent activity against MRSA has been synthesized via a Stille cross-coupling reaction. These new carbapenems show reduced serum protein binding and improved in vivo efficacy as a consequence of the positively charged amidinium substituent. © 1999 Elsevier Science Ltd. All rights reserved.

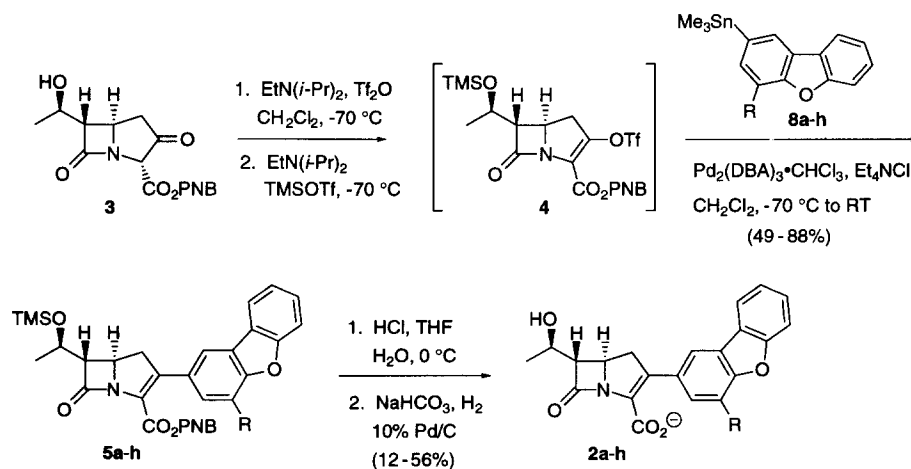
Introduction: Infections caused by methicillin-resistant *Staphylococcus aureus* (MRSA) and methicillin-resistant coagulase negative staphylococci (MRCNS) are becoming an increasingly serious clinical problem worldwide.¹ In these laboratories, there has been an ongoing effort to identify carbapenem antibiotics to combat these infections. Investigation of various 2-arylcabapenems led to the discovery of 2-*meta*-biphenyl-carbapenems with excellent activity against MRSA/MRCNS.² Increased potency was achieved by incorporating the biphenyl moiety into a planar tricyclic ring system such as a dibenzofuran, carboline, or fluorenone.³ In addition, it has been found that an electron withdrawing substituent at the 4-position of a 2-dibenzofuranylcabapenem (e.g., **1b,c**) results in a further enhancement of MRSA/MRCNS activity. However, hydrophobic analogs such as **1** show extremely high levels of binding to serum proteins and, as a result, are not highly efficacious in vivo. As a means of overcoming this problem, we have employed positively charged amidinium substituents as electronegative groups at the 4-position of the dibenzofuranyl moiety. The resulting zwitterionic carbapenems **2** have been found to be less highly bound to human plasma proteins than the net anionic derivatives **1** and in most cases retain excellent in vitro activity against MRSA and MRCNS. Improved in vivo efficacy in a mouse systemic infection model has also been demonstrated for carbapenems **2**.



Chemistry: The 2-dibenzofuranylcabapenems **2** were synthesized via a slight modification of our previously described Stille cross-coupling method (Scheme 1).⁴ The known β -ketoester **3** was converted to the corresponding enol triflate (Tf₂O, EtN(*i*-Pr)₂, CH₂Cl₂, -70 °C) followed by in situ protection of the

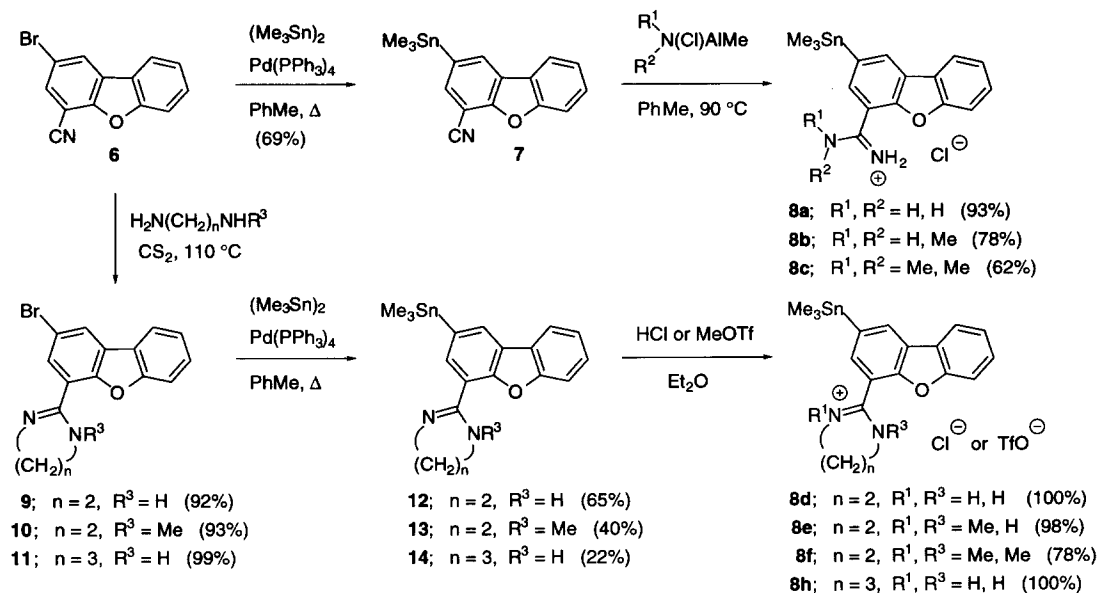
hydroxyethyl sidechain as its TMS ether [TMSOTf, EtN(*i*-Pr)₂] to give **4**. Without isolation, enol triflate **4** was reacted with the requisite aryl-stannane **8a–h** employing Pd₂(dba)₃•CHCl₃ as catalyst and Et₄NCl as a soluble chloride source (CH₂Cl₂, -70 °C to RT). After conventional aqueous work-up, coupled products **5a–h** were isolated in 49–88% yield by precipitation from CH₂Cl₂-Et₂O. Significantly, this procedure accomplished introduction of the dibenzofuran sidechain with the fully elaborated amidinium substituent in a one-pot sequence. Acidic hydrolysis of the silyl group followed by hydrogenolysis of the *p*-nitrobenzyl ester and finally reverse-phase chromatography gave the desired carbapenems **2** in 12–56% yield.

Scheme 1. Synthesis of 2-Dibenzofuranylcabapenems **2a–h** by Stille Cross-Coupling



The amidinium-stannanes **8a–h** were synthesized as shown in Scheme 2 starting with 2-bromo-4-cyano-dibenzofuran **6**.^{5,6} Stannylation of **6** in standard fashion with hexamethylditin provided **7**. Conversion of nitrile **7** to the amidines **8a–c** was accomplished by the one-step method of Garigipati,⁷ employing the appropriate aluminum amide reagent⁸ in each case. Amidines **8a–c** were isolated as hydrochloride salts after chromatography on silica gel (EtOAc, MeOH, CH₂Cl₂). The cyclic amidines were synthesized by heating nitrile **6** with a large excess of the appropriate 1,2-ethylenediamine or 1,3-propylenediamine derivative and catalytic carbon disulfide⁹ to provide **9–11**. Stannylation followed by protonation or methylation then yielded stannanes **8d–f** and **8h**. Stannane **8g** was synthesized from intermediate **10** by dehydrogenation with barium manganate followed by stannylation and methylation.^{10,11}

Biological Activity: As shown in Table 1, the simple amidine substituted compounds **2a** and **2d** retained the excellent anti-MRSA/MRCNS activity of compounds **1**. When compared to vancomycin, the therapeutic agent of choice for these pathogens, **2a** and **2d** were 1–2 times as active against MRSA and 4–8 fold more active against MRCNS. However, further substitution of the amidine resulted in a progressive loss of anti-MRS activity. This was particularly striking in the cyclic amidine series where **2e** and **2f** were 4 fold and 32 fold less active than **2d** respectively. This may reflect a requirement for co-planarity of the amidine with the aromatic ring system for optimal activity. Such a conformation would be highly disfavored in compound **2f**. Interestingly, the imidazolium analog **2g** was somewhat more active than **2f**. In addition, the 6-membered cyclic amidine compound, **2h**, was found to be 2–4 fold less active than its 5-membered counterpart **2d**.

Scheme 2. Synthesis of Amidinium Stannanes 8a–h

As expected, the amidinium substituted compounds **2a–h** were much less highly serum bound than carbapenems **1**. For example, compound **1b** was found to be 98% bound to human plasma proteins whereas **2a** and **2d** were only 71% and 34% bound respectively (Table 2). In addition, **2a** and **2d** showed little or no MIC elevation in the presence of serum, while the MICs of **1b** and **1c** were highly serum antagonized. This proved to be predictive of the in vivo performance of the compounds. In a mouse systemic infection model, against a challenge dose of *S. aureus* MB2985 (methicillin-susceptible), amidines **2a** and **2d** were both more than 10 fold more efficacious than **1b** and 4–5 times more active than **1c**.

Table 1. Anti-MRSA/MRCNS Activity^a of 1 and 2

Compd	MRSA (12)			MRCNS (9)		
	Range	MIC ₅₀	MIC ₉₀	Range	MIC ₅₀	MIC ₉₀
1a	0.5–8	2	4	0.5–8	4	8
1b	0.125–2	0.5	1	0.125–1	1	1
1c	0.25–2	0.5	1	0.25–2	2	2
2a	0.25–4	1	2	0.5–4	1	2
2b	0.5–4	1	4	0.5–2	2	2
2c	1–8	2	8	1–8	4	8
2d	0.25–2	1	1	0.25–1	1	1
2e	1–8	2	4	1–8	2	4
2f	2–32	8	32	8–32	16	32
2g	2–16	4	8	4–16	8	16
2h	0.5–4	1	2	1–4	2	4
VAN ^b	1–2	1	2	2–8	4	8
IPM ^b	1–128	32	128	32–>128	128	>128

(a) MICs ($\mu\text{g/mL}$) were determined using the broth microtube dilution method. Mueller-Hinton Broth + 2% NaCl, inoculum $\sim 10^5$ cfu/mL, incubation at 35°C for 46 h. MICs read to no visible growth (ref 2).

(b) VAN = vancomycin; IPM = imipenem. Data reflect the mode of 9 measurements of each panel of strains.

Table 2. In Vivo Efficacy of 1 and 2 in a Mouse Systemic Infection Model^a

Compd	%PB ^b	MIC ($\mu\text{g/mL}$)	Plasma MIC ^c ($\mu\text{g/mL}$)	ED ₅₀ (mg/kg)
1b	98	<0.016	>32	1.44
1c	95	0.031	1	0.55
2a	71	0.031	0.031	<0.096
2d	34	≤ 0.016	0.063	0.135
IPM (n=5)	≤ 5	0.010 (± 0.004)	0.014 (± 0.004)	0.026 (± 0.006)

(a) Charles River CD-1 female mice. Infecting organism *S. aureus* MB2985 (methicillin-susceptible). Challenge Dose 4.0×10^7 cfu IP in brain heart broth. Antibiotics administered SC at 0 h.

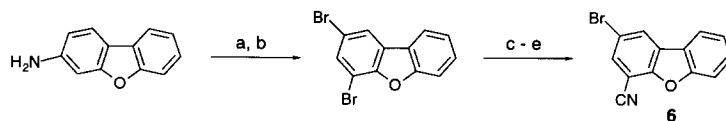
(b) Percent binding to human albumin fraction V.

(c) MIC in presence of human albumin fraction V (21.5 mg/mL).

Conclusions: Positively charged amidinium substituents were employed to reduce the serum protein binding and improve the in vivo performance of a series of 2-dibenzofuranylcabapenems, while in most cases maintaining excellent activity against MRSA and MRCNS. The most potent compounds, **2a** and **2d**, were more active than vancomycin against MRSA and MRCNS in vitro and demonstrated good in vivo efficacy in a mouse infection model. The compounds described herein thus represent a step forward in our quest to identify new carbapenem antibacterial agents for the treatment of infections due to these troublesome pathogens.

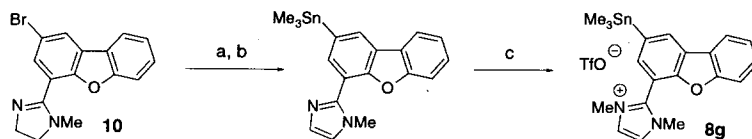
References and Notes

- (a) Voss, A.; Doebebeling, B. N. *Int. J. Antimicrob. Agents* **1995**, *5*, 101. (b) Brumfitt, W.; Hamilton-Miller, J. M. T. *Drugs Exptl. Clin. Res.* **1994**, *20*, 215. (c) Chambers, H. F. *Clin. Microbiol. Rev.* **1997**, *10*, 781. (d) Ayliffe, G. A. J. *Clin. Infect. Dis.* **1997**, *24* (Suppl 1), S74.
- DiNinno, F.; Muthard, D. A.; Salzmann, T. N.; Huber, J.; Kahan, J.; Kropp, H. *Bioorg. & Med. Chem. Lett.* **1995**, *5*, 945.
- Meurer, L. C.; Guthikonda, R. N.; Huber, J. L.; DiNinno, F. *Bioorg. & Med. Chem. Lett.* **1995**, *5*, 767.
- Rano, T. A.; Greenlee, M. L.; DiNinno, F. P. *Tetrahedron Lett.* **1990**, *31*, 2853.
- Prepared from 3-aminodibenzofuran (ref 6) in 5-steps:



(a) Br₂, dioxane, 2N NaOH, 89%; (b) *t*-BuONO, DMF, 50 °C, 68%; (c) i. *n*-BuLi, THF, -70 °C; ii. DMF, -50 °C; (d) H₂NOH • HCl, pyridine, EtOH, 100%; (e) Tf₂O, Et₃N, CH₂Cl₂, -70 °C, 77%.

- Gilman, H.; Avakian, S. *J. Am. Chem. Soc.* **1946**, *68*, 580.
- Garigipati, R. S. *Tetrahedron Lett.* **1990**, *31*, 1969.
- Levin, J. I.; Turos, E.; Weinreb, S. M. *Synth. Commun.* **1982**, *12*, 989.
- Leistner, S.; Wagner, G.; Krasselt, U.; Dumke, S. *Pharmazie* **1992**, *47*, 11.
- Synthesis of stannane **8g**:



(a) BaMnO₄, CH₂Cl₂, 45 °C, 83% (ref 11); (b) (Me₃Sn)₂, Pd(PPh₃)₄, PhMe, Δ, 83%; (c) MeOTf, CH₂Cl₂, 90%.

- Hughey, J. L.; Knapp, S.; Schugar, H. *Synthesis* **1980**, 489.